

Engineering Nanomedical Systems

Weldon School

cal Engineering

Lecture 5 Nanomaterials for core design

BME 695

James F. Leary, Ph.D.

SVM Endowed Professor of Nanomedicine Professor of Basic Medical Sciences and Biomedical Engineering

Member: Purdue Cancer Center; Oncological Sciences Center; Bindley Biosciences Center, Birck Nanotechnology Center Email: jfleary@purdue.edu

Copyright 2011 J.F. Leary

5.1 Introduction

A. core building blocks

- B. functional cores for theranostics
- C. "functionalizing" the core surface chemistry to attach other molecules

Types of Core Materials and their detection

Core material Detection Iron oxide x-ray, MRI, add fluorescent probe C60 and carbon nanotubes add fluorescent probe Gold surface plasmon resonance Silver surface plasmon resonance add fluorescent probe Silica Quantum dots intrinsic long-lifetime fluorescence "Next generation" quantum dots intrinsic fluorescence Hybrid materials mixture of detectable properties

5.2 Ferric oxide cores

- A. paramagnetic cores
- B. superparamagnetic cores
- C. ferric nanorods
- D. advantages and disadvantages

Ferric Oxide Nanospheres and Nanorods

(a) (b)



Figure 1. Transmission electron micrographs (TEM) of (a) spherical iron nanoparticles with diameters of 2 nm, and (b), rod-shaped iron nanoparticles with dimensions of 2 nm 11 nm, (Inset: High-resolution electron micrograph of a single nanorod) The images were obtained with a JEOL JEM-2000EX II instrument. From: Park et al., J. Am Chem. Soc. 2000

5.3 C60 and carbon nanotubes

- A. size and structure of C60
- B. elongation of C60 into carbon nanotubes
- C. advantages and disadvantages

C60 "Buckyballs" and Carbon nanotubes as drug carriers



http://www.ydae.edu



http://www.udel.edu

5.4 Gold cores

- A. gold nanoparticles
- B. gold nanorods
- C. other shapes (e.g. "stars")
- D. gold nanoshells
- E. advantages and disadvantages

Gold and silver nanoparticles



200nm (same for all the images)

From: Rosi and Mirkin, 2005.

Gold nanorods for optical imaging







Courtesy: Ji-Xin Cheng and Alex Wei

Gold nanorods, which fluoresce red, were photographed inside the blood vessels of a live mouse by researchers in Purdue's Weldon School of Biomedical Engineering and Department of Chemistry. The researchers have taken a step toward developing a new type of ultra-sensitive medical imaging technique that works by shining a laser through the skin to detect the tiny rods injected into the bloodstream. In tests with mice, the nanorods yielded images nearly 60 times brighter than conventional fluorescent dyes, including rhodamine, commonly used for a wide range of biological imaging to study the workings of cells and molecules. (Purdue University photo courtesy of Weldon School of Biomedical Engineering and Department of Chemistry)

5.5 Silica cores

- A. Silica nanoparticles
- B. Embedding fluorophores to prevent photobleaching
- C. Other advantages and disadvantages

Possibility of low toxicity silicon nanoparticles for in-vivo use? (Allison Hubel group and collaborators at Univ. Minnesota)



Achievements:

 Scaleable high-yield synthesis approach for silicon quantum dots

Mangolini et al., NanoLett 5, 655, 2005



Achievements:

- Organic surface passivation
- Photoluminescence quantum yield > 60%

Jurbergs et al., Appl. Phys. Lett., June 5, 2006

Fundamental issues:

- Crystal formation in low-temperature plasmas
- Surface properties of quantum dots

Source: Research Highlights, Univ. Minnesota 2006

Silica nanoparticles can be easily made in different sizes and can embed conventional fluorescent molecules and prevent photobleaching



- FIGURE 1. Transmission electron micrographs of different sizes of silica nanoparticles prepared in various microemulsion systems.
- (a) 15-nm nanoparticles; (b) 40-nm nanoparticles; (c) 120-nm nanoparticles; scale bars are 200 nm, 200 nm, and 1 μ m, respectively.

From Wang et al., 2006.

Use of fluorescent dye embedded silica nanoparticles



FIGURE 3. (a) SEM image of an E. coli O157:H7 cell incubated with nanoparticle-antibody conjugates, showing

nanoparticle binding to the target bacterium. Scale bar is 2.73 µm. (b) SEM image of an E. coli DH5a cell (negative control) incubated with nanoparticle–antibody conjugates, showing no nanoparticle binding. Scale bar is 1.5 µm. The black small dots in (a) and (b) are the pores on the surface of the filter membrane, and the white spots are unbound nanoparticles. (c) Fluorescence image of an E. coli O157:H7 cell after incubation with nanoparticle–antibody conjugates. Scale bar is 4 µm. The fluorescence intensity is strong, enabling identification of a single bacterial cell in aqueous solution.

From: Wang et al., 2006

Mesoporous Silica NPs (MSNs) for drug release



Transmission electron microscopy images of three spherical MSNs with different particle and pore sizes: a) Particle size ca. 250 nm; pore diameter ca. 2.3 nm. b) Particle size ca. 200 nm; pore diameter ca. 6.0 nm. c) Particle size ca. 50 nm; pore diameter ca. 2.7 nm.

Original source: C.-Y. Lai, B. G. Trewyn, D. M. Jeftinija, K. Jeftinija, S. Xu, S. Jeftinija, V. S. Y. Lin, *J. Am. Chem. Soc.* **2003**, *125*, 4451

5.6 Quantum Dots





* Not including coatings

Quantum Dot Nanoparticles

- Excitation/emission spectra
- Photostability
- Size tunable
- Bioconjugation





Quantum Dot Nanoparticles





Transmission electron microscopy (TEM) image of amino-functionalized Qdots. Size was determined to be ~10 nm.

Biomolecular Targeting: Peptide

- Use of biomolecules offers advantages toward other uptake mechanisms: Cell receptor is targeted and functions normally
- Peptide offers ease of synthesis and well understood chemistry. These are also on the size order of the nanoparticles.
 - QTracker® Cell Labeling Kit (Invitrogen Corporation, Carlsbad, CA) offers Qdot nanoparticles conjugated to a universal peptide. This will enter all cell lines.
 - Specific peptides will enter only certain cell types; the focus of nanomedical approach to disease





In vitro SkBr3 Study: Confocal Images



SkBr3 cells with application of Qdot-LTVSPWY complex

Quantum Dot Nanoparticles

- Agglomeration properties based on:
 - Chemistry of the Qdot; its properties, surface charge/molecules, elemental makeup, etc.
 - Chemical environment; for instance pH



Qdots imaged in biological environments. (a) *In vitro* Qdots in cell (Red, scale bar 5µm). (b) *In vivo* Qdots within tissue (White, scale bar 100µm).

In vivo SkBr3 Tumor Study: Results



Fluorescent microscopy images of *in vivo* tumor tissue.(a) Image of control kidney tissue, this sample did not receive any Qdots.

- (b) Image of tumor tissue from a peritumoral injection.
- (c) Image of tumor tissue from a tail vein injection.

Qdot Agglomeration



(a) *In vivo* tumor image. (b) Graphic representation of agglomerated Qdots.

NANOPARTICLE AGGLOMERATION:

~1000 - 2000 nm IN DIAMETER

APPROXIMATE: 50 – 100 NANOPARTICLES PER CLUSTER IN DIAMETER

CONSIDERING THREE DIMENSIONS, THE NUMBER OF NANOPARTICLES PRESENT COULD BE BETWEEN 125,000 AND 10⁶

Cytotoxicity: Some DNA damage!

- Confocal imaging
 - ROS are present normally in cells. Heightened presence indicates a state of cellular stress.
 - Detection of ROS was observed in the positive control sample and the QTracker® sample.

Dihydroethidium is shown in red QTracker® is shown in green.

(a) Control

(b) H₂O₂

(c) QTracker®



5.5 Next generation quantum dots

A. Water-Soluble Doped ZnSe Nanocrystal Emitters

B. Organic quantum dots

Future of Quantum Dots is Still being written...

- Concerns are arising over potential in-vivo toxicity of Cd based quantum dots
- But new less toxic "d-dots" are being developed



Efficient, Stable, Small, and Water-Soluble Doped ZnSe Nanocrystal Emitters as Non-Cadmium Biomedical Labels

Narayan Pradhan, David M. Battaglia, Yongcheng Liu, and Xiaogang Peng Nano Lett.; **2007**; *7*(2) pp 312 - 317; **(Letter)**

5.8 Hybrid material cores

- A. Gold-ferric oxide nanoparticles and nanorods
- B. NIR fluorescent-chitosan polymer-iron oxide core hybrids

Polymers as vehicles for SPIO NPs



work of graduate student Jaehong Key

In-vivo fluorescent imaging (eXplore Optix) of NIR (Cy5.5) fluorescent chitosan-SPIO NPs



work of graduate student Jaehong Key

Ex-Vivo imaging of NIR (Cy5.5) fluorescent chitosan-SPIO NP labeled isolated mouse organs



work of graduate student Jaehong Key

Lecture 5 References

- 1. Burda, C., Chen, X., Narayanan, R., El-Sayed, M.A. Chemistry and Properties of Nanocrystals of Different Shapes Chem. Rev. 2005, 105, 1025-1102 (a VERY comprehensive review!)
- 2. Mornet, S., Vasseur, S., Grasset, F., Duguet, E. Magnetic nanoparticle design for medical diagnosis and therapy. J. Mater. Chem 14: 2161-2175, 2004.
- Osaka, T., Matsunaga, T., Nakanishi, T., Arakaki, A., Niwa, D., Iida, H. Synthesis of magnetic nanoparticles and their application to bioassays. Anal Bioanal Chem (2006) 384: 593–600
- Park, S-J, Kim,S., Lee, S., Khim, Z.G., Char, K., Hyeon, T. Synthesis and Magnetic Studies of Uniform Iron Nanorods and Nanospheres. J. Am. Chem. Soc. 2000, 122, 8581-8582
- 5. Wang, L., Wang, K., Santra, S., Zhao, X., Hilliard, L.R., Smith, J.E., Wu, Y., Tan, W. Watching Silica Nanocrystals Glow in the Biological World. Analytical Chemistry 647-654, 2006.
- Cooper, C.L., Reece, L.M., Key, J., Bergstrom, D.E, Leary, J.F. "Water-soluble iron oxide nanoparticles for nanomedicine" a poster on nanHUB at http://docs.lib.purdue.edu/cgi/viewcontent.cgi?article=1021&context=nanoposter
- William W. Yu, Joshua C. Falkner, Cafer T. Yavuz and Vicki L. Colvin "Synthesis of monodisperse iron oxide nanocrystals by thermal decomposition of iron carboxylate salts" Chem. Commun., 2306 – 2307, 2004.
- 8. Lu, J., Liong, M., Zink, J.I., Tamanoi, F. "Mesoporous Silica Nanoparticles as a Delivery System for Hydrophobic Anticancer Drugs" Small 3(8): 1341 1346, 2007.
- 9. Igor I. Slowing, Brian G. Trewyn, Supratim Giri, and Victor S.-Y. Lin "Mesoporous Silica Nanoparticles for Drug Delivery and Biosensing Applications" Adv. Funct. Mater., *17*, 1225–1236, 2007.